# **Journal of Medical Microbiology** and Infectious Diseases

eISSN: 2345-5330

# Burkholderia gladioli Septicemia: A Rare Case Report

Priyanka Patil<sup>1\*</sup>, Mrudula Dravid<sup>1</sup>, Karuna Ahire<sup>1</sup>, Subhash Bothikar<sup>1</sup>, Farhan Shah<sup>1</sup> <sup>1</sup>JMF's ACPM Medical College, Sakri Road, Morane, Dhule, North Maharashtra, India-424002

## ARTICLE INFO

### ABSTRACT

### Case Report

**Keywords:** Burkholderia gladioli, Bacteremia, Opportunistic infection, Bacterial identification

Received: 23 Aug. 2024

Received in revised form: 30 Nov. 2024

Accepted: 15 Jan. 2025

DOI: 10.61186/JoMMID.13.2.134

### \*Correspondence

Email: priyankapatildr@gmail.com



**Introduction**: Burkholderia gladioli is an emerging opportunistic pathogen in humans. Its phenotypic variability makes identification challenging and can lead to misidentification with closely related Burkholderia species. Case **presentation**: We report a case of B. gladioli septicemia in a patient from the North Maharashtra region of India. The isolate was accurately identified using an automated bacterial identification and antimicrobial susceptibility testing system. This case highlights the need for accurate identification for appropriate management of infections caused by uncommon pathogens. Discussion: This case emphasizes the importance of considering B. gladioli the differential diagnosis, particularly in immunocompromised individuals. Accurate diagnosis and appropriate antimicrobial therapy require a high index of suspicion and advanced identification methods. This report highlights the challenges in identifying B. gladioli and emphasizes the need for increased awareness of its clinical significance and antimicrobial resistance patterns. Conclusions: B. gladioli infections, while uncommon, cause severe disease in human health, particularly in patients with potential immunocompromise or environmental exposures. Improved diagnostic capabilities, such as molecular identification, and increased awareness of B. gladioli's pathogenic potential are essential for effective management of these infections.

### INTRODUCTION

The genus Burkholderia encompasses a diverse group of Gram-negative bacilli with a wide environmental distribution. Among these, Burkholderia gladioli, originally identified as a phytopathogen affecting plants, has emerged as an opportunistic human pathogen. Although primarily a phytopathogen that affects plants such as gladiolus and onions, B. gladioli is an opportunistic human pathogen, characterized by motility, aerobic growth, oxidase positivity, and a distinctive "safety-pin" appearance on staining. Key distinguishing features include resistance to polymyxin B and colistin

B. gladioli is a common environmental pathogen found in soil, water, and various plant species, including gladiolus and decaying onions. It is well-documented as a plant pathogen [2]. While less common, B. gladioli can also cause serious human infections, including sepsis. This report highlights the need for increased awareness and further research.

B. gladioli can infect humans through various routes, potentially including ingestion and inhalation. In rare regional outbreaks, ingestion of rice contaminated with toxin-producing B. gladioli has caused foodborne illness.

Respiratory infection may result from inhaling aerosolized bacteria or contaminated soil particles. B. gladioli pathovar cocovenenans has been implicated in food poisoning outbreaks, particularly in China [3], due to the production of toxins such as bongkrekic acid and toxoflavin. Like other Gram-negative bacilli, B. gladioli produces lipopolysaccharides (LPS) and proteases; however, their specific role in human pathogenesis remains poorly defined and warrants further investigation. These toxins can cause a range of illnesses, from mild gastrointestinal symptoms to life-threatening sepsis [3].

The first reported cases of respiratory tract infections caused by B. gladioli date back to 1989, specifically in patients with cystic fibrosis (CF), a genetic disorder associated with chronic respiratory infections [4-7]. These early reports demonstrated the potential for B. gladioli to infect vulnerable individuals, especially those with underlying respiratory conditions. B. gladioli can contaminate aqueous solutions, contributing to outbreaks linked to contaminated saline solutions in healthcare settings [8, 9]. This underscores the critical importance of rigorous sterilization of medical equipment and solutions and infection control procedures in healthcare settings.

Patil et al.

B. gladioli infections have been reported in patients with cystic fibrosis, a condition associated with a compromised immune system. This risk is amplified in lung transplant recipients, as both the underlying disease and the procedure, along with the presence of indwelling medical devices, increase susceptibility to opportunistic infections [10-13]. Additionally, individuals with chronic granulomatous disease and other immunocompromising conditions, such as human immunodeficiency virus (HIV) infection, cancer, and immunosuppressive therapy, are also susceptible to B. gladioli infections [14, 15]. In these populations, infections are more likely to be severe and disseminated, necessitating prompt diagnosis and treatment.

This report describes a case of septicemia caused by *B. gladioli* in the North Maharashtra region of India. This is a rare documented case of *B. gladioli* bacteremia in this region. This case highlights the potential for *B. gladioli* to cause septicemia in this region and emphasizes the need for clinicians to consider this pathogen in patients presenting with fever, respiratory distress, or sepsis. Reporting rare cases like this contributes to a better understanding of the epidemiology and clinical presentation of *B. gladioli* infections, which can aid in earlier diagnosis and treatment.

### **CASE PRESENTATION**

A 60-year-old female resident of Nyahalod, located in the Dhule district of the North Maharashtra region, presented to the hospital on Jun 19, 2024, with complaints of a high fever (temperature 39.5°C) and difficulty breathing. Written informed consent for treatment and publication of this case report was obtained from the patient and her family. She was an onion field worker, potentially exposed to *B. gladioli* through contact with contaminated soil or decaying onions. She reported a history of recurrent chest infections with a persistent cough and thick, tenacious mucus.

On examination, her blood pressure was 80/52 mmHg (hypotensive), and oxygen saturation was 70% on room air (indicating hypoxemia, measured by pulse oximetry). Lung auscultation revealed reduced breath sounds and bilateral basal crepitations. Her respiratory rate was 26 breaths/min, and heart rate was 110 beats/min. Her white blood cell count was elevated at 26,400/µL. Chest radiography demonstrated bronchial wall thickening, consistent with chronic respiratory infection. Given her presentation, she was treated empirically for sepsis, with chronic respiratory disease as a possible underlying condition requiring further investigation. The patient was admitted to the medical ward of ACPM Medical College, Dhule, and initiated on empiric antimicrobial therapy.

Blood cultures were sent to the Microbiology Department. After 3 days of incubation in the BACTEC system, the blood cultures flagged positive for microbial growth. The colonies were non-pigmented, slightly mucoid, and non-lactose fermenting on MacConkey agar,

resembling other non-fermenters. Gram staining of the colonies revealed Gram-negative bacilli (Fig.1). The isolate was oxidase-positive, and further identification and antimicrobial susceptibility testing were performed using the VITEK 2 system. The isolate was identified as *B. gladioli*. Antimicrobial susceptibility testing revealed the isolate to be susceptible to several antibiotics, including cefoperazone/sulbactam, imipenem, meropenem, and aminoglycosides.

The patient's condition rapidly deteriorated, with worsening hypoxia and multi-organ dysfunction. She died two days after admission, before the final culture and sensitivity report was available. Empirical treatment with vancomycin and polymyxin B was initiated, targeting a broad range of suspected pathogens. However, the patient died before susceptibility results were available, which later confirmed the isolate's susceptibility to cefoperazone/sulbactam, imipenem, meropenem, and aminoglycosides.

### DISCUSSION

Accurate identification of B. gladioli in clinical microbiology laboratories can be Differentiating it from members of the Burkholderia cepacia complex (BCC) is challenging, as conventional phenotypic methods and some automated identification systems may not resolve subtle biochemical differences. This difficulty in identification likely contributes to the underreporting of B. gladioli infections, as highlighted by Boyanton et al. (2005) [14]. The limitations of current phenotypic and some automated identification methods underscore the importance of molecular techniques, such as gene sequencing, for accurate identification of B. gladioli and differentiation from closely related species within the BCC. Molecular methods offer advantages in terms of specificity, sensitivity, and the ability to detect and characterize resistance mechanisms.

In this case, *B. gladioli* was isolated from blood culture using the BACTEC system and subsequently identified using the VITEK 2 system. While blood culture remains essential for detecting bacteremia, accurate identification of rare or emerging pathogens like *B. gladioli* often requires advanced methods. Automated systems with updated databases and broader identification capabilities, particularly those incorporating molecular techniques, can facilitate this process. Rapid and accurate identification, particularly using molecular methods or updated databases in automated systems, is crucial for timely and appropriate treatment of *B. gladioli* infections, which can rapidly progress, as illustrated in this case.

B. gladioli is a rare cause of sepsis. A literature review revealed only a limited number of reported cases of B. gladioli bacteremia [10, 16, 17]. The rarity of B. gladioli septicemia can hinder prompt diagnosis and treatment, as clinicians may be unfamiliar with its clinical presentation and antimicrobial susceptibility profile. B. gladioli, primarily known as a plant pathogen, was not commonly recognized as a human pathogen until recent decades. Early studies did not implicate B. gladioli in human disease [5, 10, 18]. Reported cases of B. gladioli infection have occurred in

both adults and children, although the precise age distribution remains unclear [5, 10, 13, 14, 17, 18]. This lack of clarity regarding the age distribution, combined with limitations in identification methods and potential underreporting, likely obscures the true incidence of *B. gladioli* infections.

A 22-year-old man with cystic fibrosis and declining pulmonary function who underwent lung transplantation developed *B. gladioli* septicemia and empyema, as reported by Khan *et al.* (1996) [9]. This case highlights the potential for *B. gladioli* to cause severe infections even in patients without classical immunodeficiencies, such as organ transplant recipients or those with HIV. Dursun *et al.* (2012), in a study of neonatal septicemia, reported cases of B. gladioli bacteremia, with a 21.4% mortality rate in cases complicated by sepsis [19]. This low incidence of fever, a hallmark of bacteremia, warrants further investigation and may reflect specific characteristics of *B. gladioli* infections in neonates or limitations in the study's detection methods.

Boyanton et al. (2005) tested the susceptibility of B. gladioli isolates to various antibiotics and found them susceptible to gentamicin, amikacin, ticarcillinclavulanate, and ciprofloxacin, among others [14]. Therefore, these antibiotics represent potential treatment options, although susceptibility testing is essential to guide therapy for individual B. gladioli infections. Graves et al. (1997) found B. gladioli strains to be susceptible to range of antibiotics, including quinolones, aminoglycosides, and imipenem [10]. These antibiotic classes are thus potential treatment options for B. gladioli infections; however, individual susceptibility testing is essential. Antimicrobial susceptibility patterns of B. gladioli can vary between strains and over time, highlighting the importance of ongoing surveillance to guide optimal therapy. In the present case, antimicrobial susceptibility testing revealed that the B. gladioli isolate was susceptible to amikacin, gentamicin, imipenem, trimethoprim-sulfamethoxazole, ciprofloxacin. ceftriaxone, cefoperazone/sulbactam, and meropenem, but resistant to aztreonam, ceftazidime, and colistin. The patient was empirically treated with vancomycin and polymyxin B, but susceptibility testing later confirmed the isolate's susceptibility to cefoperazone/sulbactam and amikacin, which were not administered due to the patient's death. Combination therapy is sometimes used for serious B. gladioli infections to enhance efficacy and potentially prevent the emergence of resistance, though evidence is limited [19].

In this case report, the isolate of *B. gladioli* exhibited susceptibility to cefoperazone/sulbactam, imipenem, meropenem, amikacin, gentamicin, ciprofloxacin, and trimethoprim/sulfamethoxazole. The patient was treated with cefoperazone/sulbactam and amikacin based on antimicrobial susceptibility testing. The findings of this case emphasize the importance of clinicians considering and accurately identifying rare pathogens such as *B*.

gladioli and their antimicrobial susceptibility patterns. This knowledge is essential for guiding treatment decisions and improving patient outcomes, as prompt and appropriate antimicrobial therapy is critical for patient survival in cases of severe infection. It is also crucial to develop and update local and national guidelines to reflect emerging resistance patterns and new treatment options.

A previous study of *B. gladioli* infections in newborns reported a 21.4% in-hospital mortality rate for cases complicated by sepsis, compared to 7% for *B. gladioli* infections without sepsis [19]. In the present case, the patient's clinical course was consistent with sepsis and multiple organ failure, leading to her death. This case underscores the rapid progression and potentially fatal outcome of *B. gladioli* septicemia. Further research is needed to better understand the risk factors for severe *B. gladioli* infections, the mechanisms underlying its pathogenesis, and the optimal treatment strategies, particularly for patients with pre-existing respiratory conditions.

Given the potential for *B. gladioli* to cause hospital-acquired infections, it is imperative that standardized infection control protocols be followed to prevent transmission and reduce the risk of infection. Moreover, as *B. gladioli* can be acquired from environmental sources, environmental hygiene and infection control measures are essential for preventing infection. This includes hand hygiene according to WHO guidelines, avoiding contact with contaminated surfaces and objects, and maintaining aseptic environments and adhering to infection prevention protocols. Healthcare facilities should implement comprehensive infection prevention strategies, particularly for immunocompromised patients, those with chronic respiratory conditions, and post-transplant recipients.

This case highlights that B. gladioli, though a rare pathogen, can cause rapidly progressive and lifethreatening particularly infections, immunocompromised individuals. A limitation of this study is the reliance on phenotypic methods for bacterial identification. Molecular methods, such as 16S rRNA gene sequencing, would provide more definitive identification and could help differentiate B. gladioli from closely related species. Trimethoprim-sulfamethoxazole, commonly used for Burkholderia infections [20], was effective against the B. gladioli isolate in this case, highlighting the importance of susceptibility testing. Susceptibility testing is crucial to guide appropriate antibiotic selection. Further research is needed to better understand the clinical spectrum of B. gladioli infections and to identify optimal treatment strategies for specific patient populations, particularly those with underlyingrespiratory conditions.

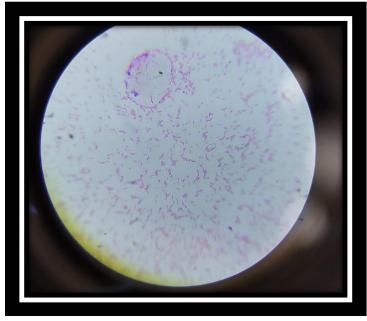
In conclusion, this case underscores the importance of considering rare pathogens like *B. gladioli* in the differential diagnosis of sepsis, especially in patients with predisposing factors. Rapid identification and appropriate

Patil et al.

antimicrobial therapy are essential for improving outcomes in these potentially life-threatening infections.

**Ethical considerations.** This case report was approved by the Institutional Ethics Committee (IEC) of JMF's

ACPM Medical College and Hospital, Dhule (IEC registration number: ECR/1448/ Inst/MH/2020; approval number: 148 IEC/ACPMMC/Dhule, dated March 4, 2024).



**Fig. 1.** Gram stain of *B. gladioli* isolated from a blood culture sample, demonstrating the bipolar ('safety pin') staining typical of *B. gladioli* (light microscopy, ×1000 magnification).

### CONFLICTS OF INTEREST

The authors declare that there are no conflicts of interest associated with this manuscript.

## **FUNDING**

No funding was received for this manuscript.

## REFERENCES

- 1. Sastry AS. Essentials of Medical Microbiology. 4th ed. Jaypee Brothers Medical Publishers Pvt Ltd; 2023. p. 534-9.
- Palleroni NJ, Genus I. Pseudomonas Migula 1894, 237AL. In: Krieg NR, Holt JG, editors. Bergey's manual of systematic bacteriology. Vol. 1. Baltimore: Williams & Wilkins; 1984. p. 141-9.
- 3. Jiao Z, Kawamura Y, Mishima N, Yang R, Li N, Liu X, et al. Need to differentiate lethal toxin-producing strains of *Burkholderia gladioli*, which cause severe food poisoning: description of *B. gladioli* pathovar *cocovenenans* and an emended description of *B. gladioli*. Microbiol Immunol. 2003; 47 (12): 915-25.
- 4. Barker PM, Wood RE, Gilligan PH. Lung infection with *Burkholderia gladioli* in a child with cystic fibrosis: acute clinical and spirometric deterioration. Pediatr Pulmonol. 1997; 23 (2): 123-5.
- Christenson JC, Welch DF, Mukwaya G, Muszynski MJ, Weaver RE, Brenner DJ. Recovery of *Pseudomonas gladioli* from respiratory tract specimens of patients with cystic fibrosis. J Clin Microbiol. 1989; 27 (2): 270-3.

- Clode FE, Metherell LA, Pitt TL. Nosocomial acquisition of Burkholderia gladioli in patients with cystic fibrosis. Am J Respir Crit Care Med. 1999; 160 (1): 374-5.
- Ferroni A, Sermet-Gaudelus I, Abachin E, Quesne G, Lenoir G, Berche P, et al. Use of 16S rRNA gene sequencing for identification of nonfermenting gram-negative bacilli recovered from patients attending a single cystic fibrosis center. J Clin Microbiol. 2002; 40 (10): 3793-7.
- Dobrović K, Mareković I, Payerl-Pal M, Andrijašević N, Škrobo T, Košćak V, et al. Outbreak of healthcare-associated bacteremia caused by *Burkholderia gladioli* due to contaminated multidose vials with saline solutions in three Croatian hospitals. Int J Infect Dis. 2022; 121: 152-6
- Khan SU, Gordon SM, Stillwell PC, Kirby TJ, Arroliga AC. Empyema and bloodstream infection caused by *Burkholderia* gladioli in a patient with cystic fibrosis after lung transplantation. Pediatr Infect Dis J. 1996; 15 (7): 637-9.
- Graves M, Robin T, Chipman AM, Wong J, Khashe S, Janda JM. Four additional cases of *Burkholderia gladioli* infection with microbiological correlates and review. Clin Infect Dis. 1997; 25 (4): 838-42.
- Landry ML, Jorgensen JH, Carroll KC, Pfaller MA, Warnock DW, Richter SS, et al. Manual of clinical microbiology. 11th ed. Washington, DC: ASM Press; 2015.
- Kennedy MP, Coakley RD, Donaldson SH, Aris RM, Hohneker K, Wedd JP, et al. *Burkholderia gladioli*: five year experience in a cystic fibrosis and lung transplantation center. J Cyst Fibros. 2007; 6 (4): 267-73.

- Segonds C, Clavel-Batut P, Thouverez M, Grenet D, Coustumier AL, Plésiat P, et al. Microbiological and epidemiological features of clinical respiratory isolates of *Burkholderia gladioli*. J Clin Microbiol. 2009; 47 (5): 1510-6.
- 14. Boyanton Jr BL, Noroski LM, Reddy H, Dishop MK, Hicks MJ, Versalovic J, et al. *Burkholderia gladioli* osteomyelitis in association with chronic granulomatous disease: case report and review. Pediatr Infect Dis J. 2005; 24 (9): 837-9.
- Imataki O, Kita N, Nakayama-Imaohji H, Kida JI, Kuwahara T, Uemura M. Bronchiolitis and bacteraemia caused by Burkholderia gladioli in a non-lung transplantation patient. New Microbes New Infect. 2014; 2 (6): 175-6.
- 16. Hoare S, Cant AJ. Chronic granulomatous disease presenting as severe sepsis due to *Burkholderia gladioli*. Clin Infect Dis. 1996; 23 (2): 411.

- 17. Shin JH, Kim SH, Shin MG, Suh SP, Ryang DW, Jeong MH. Bacteremia due to *Burkholderia gladioli*: case report. Clin Infect Dis. 1997; 25 (5): 1264-5.
- 18. Ross JP, Holland SM, Gill VJ, DeCarlo ES, Gallin GI. Severe *Burkholderia* (*Pseudomonas*) *gladioli* infection in chronic granulomatous disease: report of two successfully treated cases. Clin Infect Dis. 1995; 21 (5): 1291-3.
- Dursun A, Zenciroglu A, Karagol BS, Hakan N, Okumus N, Gol N, et al. *Burkholderia gladioli* sepsis in newborns. Eur J Pediatr. 2012; 171 (10): 1503-9.
- Avgeri SG, Matthaiou DK, Dimopoulos G, Grammatikos AP, Falagas ME. Therapeutic options for *Burkholderia cepacia* infections beyond co-trimoxazole: a systematic review of the clinical evidence. Int J Antimicrob Agents. 2009; 33 (5): 394-404.

Cite	this	article:
CILL	uns	ai ucic.

Patil P, Dravid M, Ahire K, Bothikar S, Shah F. *Burkholderia gladioli* Septicemia: A Rare Case Report. J Med Microbiol Infect Dis, 2025; 13 (2): 134-138. DOI: 10.61186/JoMMID.13.2.134.